

**METHOD 515.1**

**DETERMINATION OF CHLORINATED ACIDS IN WATER BY GAS  
CHROMATOGRAPHY WITH AN ELECTRON CAPTURE DETECTOR**

**Revision 4.0**

R.C. Dressman and J.J. Lichtenberg -- EPA 600/4-81-053, Revision 1.0 (1981)

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Method 3, Revision 3.0 (1987)

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## METHOD 515.1

### DETERMINATION OF CHLORINATED ACIDS IN WATER BY GAS CHROMATOGRAPHY WITH AN ELECTRON CAPTURE DETECTOR

#### 1.0 SCOPE AND APPLICATION

- 1.1 This is a gas chromatographic (GC) method applicable to the determination of certain chlorinated acids in ground water and finished drinking water.<sup>1</sup> The following compounds can be determined by this method:

Analyte	Chemical Abstract Services Registry Number
Acifluorfen*	50594-66-6
Bentazon	25057-89-0
Chloramben*	133-90-4
2,4-D	94-75-7
Dalapon*	75-99-0
2,4-DB	94-82-6
DCPA acid metabolites <sup>a</sup>	
Dicamba	1918-00-9
3,5-Dichlorobenzoic acid	51-36-5
Dichlorprop	120-36-5
Dinoseb	88-85-7
5-Hydroxydicamba	7600-50-2
4-Nitrophenol*	100-02-7
Pentachlorophenol (PCP)	87-86-5
Picloram	1918-02-1
2,4,5-T	93-76-5
2,4,5-TP	93-72-1

<sup>a</sup>DCPA monoacid and diacid metabolites included in method scope; DCPA diacid metabolite used for validation studies.

\*These compounds are only qualitatively identified in the National Pesticides Survey (NPS) Program. These compounds are not quantitated because control over precision has not been accomplished.

- 1.2 This method may be applicable to the determination of salts and esters of analyte acids. The form of each acid is not distinguished by this method. Results are calculated and reported for each listed analyte as the total free acid.
- 1.3 This method has been validated in a single laboratory and estimated detection limits (EDLs) have been determined for the analytes above (Section 13.0). Observed detection limits may vary between ground waters, depending upon the nature of interferences in the sample matrix and the specific instrumentation used.

- 1.4 This method is restricted to use by or under the supervision of analysts experienced in the use of GC and in the interpretation of gas chromatograms. Each analyst must demonstrate the ability to generate acceptable results with this method using the procedure described in Section 10.3.
- 1.5 Analytes that are not separated chromatographically, i.e., which have very similar retention times, cannot be individually identified and measured in the same calibration mixture or water sample unless an alternate technique for identification and quantitation exist (Section 11.8).
- 1.6 When this method is used to analyze unfamiliar samples for any or all of the analytes above, analyte identifications must be confirmed by at least one additional qualitative technique.

## **2.0 SUMMARY OF METHOD**

- 2.1 A measured volume of sample of approximately 1 L is adjusted to pH 12 with 6 N sodium hydroxide and shaken for one hour to hydrolyze derivatives. Extraneous organic material is removed by a solvent wash. The sample is acidified, and the chlorinated acids are extracted with ethyl ether by shaking in a separatory funnel or mechanical tumbling in a bottle. The acids are converted to their methyl esters using diazomethane as the derivatizing agent. Excess derivatizing reagent is removed, and the esters are determined by capillary column/GC using an electron capture detector (ECD).
- 2.2 The method provides a Florisil cleanup procedure to aid in the elimination of interferences that may be encountered.

## **3.0 DEFINITIONS**

- 3.1 Internal Standard -- A pure analyte(s) added to a solution in known amount(s) and used to measure the relative responses of other method analytes and surrogates that are components of the same solution. The internal standard must be an analyte that is not a sample component.
- 3.2 Surrogate Analyte -- A pure analyte(s), which is extremely unlikely to be found in any sample, and which is added to a sample aliquot in known amount(s) before extraction and is measured with the same procedures used to measure other sample components. The purpose of a surrogate analyte is to monitor method performance with each sample.
- 3.3 Laboratory Duplicates (LD1 and LD2) -- Two sample aliquots taken in the analytical laboratory and analyzed separately with identical procedures. Analyses of LD1 and LD2 give a measure of the precision associated with laboratory procedures, but not with sample collection, preservation, or storage procedures.
- 3.4 Field Duplicates (FD1 and FD2) -- Two separate samples collected at the same time and place under identical circumstances and treated exactly the same

throughout field and laboratory procedures. Analyses of FD1 and FD2 give a measure of the precision associated with sample collection, preservation and storage, as well as with laboratory procedures.

- 3.5 Laboratory Reagent Blank (LRB) -- An aliquot of reagent water that is treated exactly as a sample including exposure to all glassware, equipment, solvents, reagents, internal standards, and surrogates that are used with other samples. The LRB is used to determine if method analytes or other interferences are present in the laboratory environment, the reagents, or the apparatus.
- 3.6 Field Reagent Blank (FRB) -- Reagent water placed in a sample container in the laboratory and treated as a sample in all respects, including exposure to sampling site conditions, storage, preservation and all analytical procedures. The purpose of the FRB is to determine if method analytes or other interferences are present in the field environment.
- 3.7 Laboratory Performance Check Solution (LPC) -- A solution of method analytes, surrogate compounds, and internal standards used to evaluate the performance of the instrument system with respect to a defined set of method criteria.
- 3.8 Laboratory Fortified Blank (LFB) -- An aliquot of reagent water to which known quantities of the method analytes are added in the laboratory. The LFB is analyzed exactly like a sample, and its purpose is to determine whether the methodology is in control, and whether the laboratory is capable of making accurate and precise measurements at the required method detection limit.
- 3.9 Laboratory Fortified Sample Matrix (LFM) -- An aliquot of an environmental sample to which known quantities of the method analytes are added in the laboratory. The LFM is analyzed exactly like a sample, and its purpose is to determine whether the sample matrix contributes bias to the analytical results. The background concentrations of the analytes in the sample matrix must be determined in a separate aliquot and the measured values in the LFM corrected for background concentrations.
- 3.10 Stock Standard Solution -- A concentrated solution containing a single certified standard that is a method analyte, or a concentrated solution of a single analyte prepared in the laboratory with an assayed reference compound. Stock standard solutions are used to prepare primary dilution standards.
- 3.11 Primary Dilution Standard Solution -- A solution of several analytes prepared in the laboratory from stock standard solutions and diluted as needed to prepare calibration solutions and other needed analyte solutions.
- 3.12 Calibration Standard (CAL) -- A solution prepared from the primary dilution standard solution and stock standard solutions of the internal standards and surrogate analytes. The CAL solutions are used to calibrate the instrument response with respect to analyte concentration.

- 3.13 Quality Control Sample (QCS) -- A sample matrix containing method analytes or a solution of method analytes in a water miscible solvent which is used to fortify reagent water or environmental samples. The QCS is obtained from a source external to the laboratory, and is used to check laboratory performance with externally prepared test materials.

#### 4.0 INTERFERENCES

- 4.1 Method interferences may be caused by contaminants in solvents, reagents, glassware and other sample processing apparatus that lead to discrete artifacts or elevated baselines in gas chromatograms. All reagents and apparatus must be routinely demonstrated to be free from interferences under the conditions of the analysis by running laboratory reagent blanks as described in Section 10.2.

4.1.1 Glassware must be scrupulously cleaned.<sup>2</sup> Clean all glassware as soon as possible after use by thoroughly rinsing with the last solvent used in it. Follow by washing with hot water and detergent and thorough rinsing with dilute acid, tap and reagent water. Drain dry, and heat in an oven or muffle furnace at 400°C for one hour. Do not heat volumetric ware. Thermally stable materials such as PCBs might not be eliminated by this treatment. Thorough rinsing with acetone may be substituted for the heating. After drying and cooling, seal and store glassware in a clean environment to prevent any accumulation of dust or other contaminants. Store inverted or capped with aluminum foil.

4.1.2 The use of high purity reagents and solvents helps to minimize interference problems. Purification of solvents by distillation in all-glass systems may be required.

WARNING: When a solvent is purified, stabilizers added by the manufacturer are removed, thus potentially making the solvent hazardous. Also, when a solvent is purified, preservatives added by the manufacturer are removed, thus potentially reducing the shelf-life.

- 4.2 The acid forms of the analytes are strong organic acids which react readily with alkaline substances and can be lost during sample preparation. Glassware and glass wool must be acid-rinsed with 1 N hydrochloric acid and the sodium sulfate must be acidified with sulfuric acid prior to use to avoid analyte losses due to adsorption.
- 4.3 Organic acids and phenols, especially chlorinated compounds, cause the most direct interference with the determination. Alkaline hydrolysis and subsequent extraction of the basic sample removes many chlorinated hydrocarbons and phthalate esters that might otherwise interfere with the electron capture analysis.
- 4.4 Interferences by phthalate esters can pose a major problem in pesticide analysis when using the ECD. These compounds generally appear in the chromatogram as large peaks. Common flexible plastics contain varying amounts of phthalates,

that are easily extracted or leached during laboratory operations. Cross contamination of clean glassware routinely occurs when plastics are handled during extraction steps, especially when solvent-wetted surfaces are handled. Interferences from phthalates can best be minimized by avoiding the use of plastics in the laboratory. Exhaustive purification of reagents and glassware may be required to eliminate background phthalate contamination.<sup>3,4</sup>

- 4.5 Interfering contamination may occur when a sample containing low concentrations of analytes is analyzed immediately following a sample containing relatively high concentrations of analytes. Between-sample rinsing of the sample syringe and associated equipment with methyl-t-butyl-ether (MTBE) can minimize sample cross contamination. After analysis of a sample containing high concentrations of analytes, one or more injections of MTBE should be made to ensure that accurate values are obtained for the next sample.
- 4.6 Matrix interferences may be caused by contaminants that are coextracted from the sample. Also, note that all analytes listed in the Scope and Application section are not resolved from each other on any one column, i.e., one analyte of interest may be an interferant for another analyte of interest. The extent of matrix interferences will vary considerably from source to source, depending upon the water sampled. The procedures in Section 11.0 can be used to overcome many of these interferences. Positive identifications should be confirmed (Section 11.8).
- 4.7 It is important that samples and working standards be contained in the same solvent. The solvent for working standards must be the same as the final solvent used in sample preparation. If this is not the case, chromatographic comparability of standards to sample may be affected.

## 5.0 SAFETY

- 5.1 The toxicity or carcinogenicity of each reagent used in this method has not been precisely defined; however, each chemical compound must be treated as a potential health hazard. Accordingly, exposure to these chemicals must be reduced to the lowest possible level. The laboratory is responsible for maintaining a current awareness file of OSHA regulations regarding the safe handling of the chemicals specified in this method. A reference file of material safety data sheets should also be made available to all personnel involved in the chemical analysis. Additional references to laboratory safety are available and have been identified<sup>6-8</sup> for the information of the analyst.
- 5.2 Diazomethane -- A toxic carcinogen which can explode under certain conditions. The following precautions must be followed:
  - 5.2.1 Use only a well ventilated hood -- do not breath vapors.
  - 5.2.2 Use a safety screen.
  - 5.2.3 Use mechanical pipetting aides.

- 5.2.4 Do not heat above 90°C -- **EXPLOSION** may result.
  - 5.2.5 Avoid grinding surfaces, ground glass joints, sleeve bearings, glass stirrers -- **EXPLOSION** may result.
  - 5.2.6 Store away from alkali metals -- **EXPLOSION** may result.
  - 5.2.7 Solutions of diazomethane decompose rapidly in the presence of solid materials such as copper powder, calcium chloride, and boiling chips.
  - 5.2.8 The diazomethane generation apparatus used in the esterification procedures (Sections 11.4 and 11.5) produces micromolar amounts of diazomethane to minimize safety hazards.
- 5.3 Ethyl Ether -- Nanograde, redistilled in glass, if necessary.
- 5.3.1 Ethyl ether is an extremely flammable solvent. If a mechanical device is used for sample extraction, the device should be equipped with an explosion-proof motor and placed in a hood to avoid possible damage and injury due to an explosion.
  - 5.3.2 Must be free of peroxides as indicated by EM Quant test strips (available from Scientific Products Co., Cat. No. P1126-8, and other suppliers).

WARNING: When a solvent is purified, stabilizers added by the manufacturer are removed, thus potentially making the solvent hazardous.

**6.0** **APPARATUS AND EQUIPMENT** (All specifications are suggested. Catalog numbers are included for illustration only.)

- 6.1 Sample Bottle -- Borosilicate, 1 L volume with graduations (Wheaton Media/Lab bottle 219820 or equivalent), fitted with screw caps lined with TFE-fluorocarbon. Protect samples from light. The container must be washed and dried as described in Section 4.1.1 before use to minimize contamination. Cap liners are cut to fit from sheets (Pierce Catalog No. 012736) and extracted with methanol overnight prior to use.
- 6.2 Glassware
  - 6.2.1 Separatory funnel -- 2000 mL, with TFE-fluorocarbon stopcocks, ground glass or TFE-fluorocarbon stoppers.
  - 6.2.2 Tumbler bottle -- 1.7 L (Wheaton Roller Culture Vessel or equivalent), with TFE-fluorocarbon lined screw cap. Cap liners are cut to fit from sheets (Pierce Catalog No. 012736) and extracted with methanol overnight prior to use.

- 6.2.3 Concentrator tube, Kuderna-Danish (K-D) -- 10 mL or 25 mL, graduated (Kontes K-570050-2525 or Kontes K-570050-1025 or equivalent). Calibration must be checked at the volumes employed in the test. Ground glass stoppers are used to prevent evaporation of extracts.
- 6.2.4 Evaporative flask, K-D -- 500 mL (Kontes K-570001-0500 or equivalent). Attach to concentrator tube with springs.
- 6.2.5 Snyder column, K-D -- Three-ball macro (Kontes K-503000-0121 or equivalent).
- 6.2.6 Snyder column, K-D -- Two-ball micro (Kontes K-569001-0219 or equivalent).
- 6.2.7 Flask, round-bottom -- 500 mL with 24/40 ground glass joint.
- 6.2.8 Vials -- glass, 5-10 mL capacity with TFE-fluorocarbon lined screw cap.
- 6.2.9 Disposable pipets -- sterile plugged borosilicate glass, 5 mL capacity (Corning 7078-5N or equivalent).
- 6.3 Separatory Funnel Shaker -- Capable of holding 2 L separatory funnels and shaking them with rocking motion to achieve thorough mixing of separatory funnel contents (available from Eberbach Co. in Ann Arbor, MI or other suppliers).
- 6.4 Tumbler -- Capable of holding tumbler bottles and tumbling them end-over-end at 30 turns/min (Associated Design and Mfg. Co., Alexandria, VA and other suppliers).
- 6.5 Boiling Stones -- Teflon, Chemware (Norton Performance Plastics No. 015021 and other suppliers).
- 6.6 Water Bath -- Heated, capable of temperature control ( $\pm 2^{\circ}\text{C}$ ). The bath should be used in a hood.
- 6.7 Balance -- Analytical, capable of accurately weighing to the nearest 0.0001 g.
- 6.8 Diazomethane Generator -- Assemble from two 20 x 150 mm test tubes, two Neoprene rubber stoppers, and a source of nitrogen as shown in Figure 1 (available from Aldrich Chemical Co.). When esterification is performed using diazomethane solution, the diazomethane collector is cooled in an approximately 2 L thermos for ice bath or a cryogenically cooled vessel (Thermoelectrics Unlimited Model SK-12 or equivalent).
- 6.9 Glass Wool -- Acid washed (Supelco 2-0383 or equivalent) and heated at  $450^{\circ}\text{C}$  for four hours.

- 6.10 Gas Chromatograph -- Analytical system complete with temperature programmable GC suitable for use with capillary columns and all required accessories including syringes, analytical columns, gases, detector and stripchart recorder. A data system is recommended for measuring peak areas. Table 1 lists retention times observed for method analytes using the columns and analytical conditions described below.
- 6.10.1 Column 1 (Primary column) -- 30 m long x 0.25 mm I.D. DB-5 bonded fused silica column, 0.25  $\mu\text{m}$  film thickness (J&W Scientific). Helium carrier gas flow is established at 30 cm/sec linear velocity and oven temperature is programmed from 60-300°C at 4°C/min. Data presented in this method were obtained using this column. The injection volume was 2  $\mu\text{L}$  splitless mode with 45 second delay. The injector temperature was 250°C and the detector was 320°C. Alternative columns may be used in accordance with the provisions described in Section 10.2.
- 6.10.2 Column 2 (Confirmation column) -- 30 m long x 0.25 mm I.D. DB-1701 bonded fused silica column, 0.25  $\mu\text{m}$  film thickness (J&W Scientific). Helium carrier gas flow is established at 30 cm/sec linear velocity and oven temperature is programmed from 60-300°C at 4°C/min.
- 6.10.3 Detector -- Electron capture. This detector has proven effective in the analysis of fortified reagent and artificial ground waters. An ECD was used to generate the validation data presented in this method. Alternative detectors, including a mass spectrometer, may be used in accordance with the provisions described in Section 10.3.

## 7.0 REAGENTS AND CONSUMABLE MATERIALS

WARNING: When a solvent is purified, stabilizers added by the manufacturer are removed, thus potentially making the solvent hazardous. Also, when a solvent is purified, preservatives added by the manufacturer are removed, thus potentially reducing the shelf-life.

- 7.1 Acetone, Methanol, Methylene Chloride, MTBE -- Pesticide quality or equivalent.
- 7.2 Ethyl Ether -- Unpreserved, Nanograde, redistilled in glass if necessary. Must be free of peroxides as indicated by EM Quant test strips (available from Scientific Products Co., Cat. No. PI126-8, and other suppliers). Procedures recommended for removal of peroxides are provided with the test strips.
- 7.3 Sodium Sulfate -- Granular, anhydrous, ACS grade. Heat treat in a shallow tray at 450°C for a minimum of four hours to remove interfering organic substances. Acidify by slurring 100 g sodium sulfate with enough ethyl ether to just cover the solid. Add 0.1 mL concentrated sulfuric acid and mix thoroughly. Remove the ether under vacuum. Mix 1 g of the resulting solid with 5 mL of reagent water and measure the pH of the mixture. The pH must be below pH 4. Store at 130°C.

- 7.4 Sodium Thiosulfate -- Granular, anhydrous, ACS grade.
- 7.5 Sodium Hydroxide (NaOH) -- Pellets, ACS grade.
  - 7.5.1 NaOH, 6 N -- Dissolve 216 g NaOH in 900 mL reagent water.
- 7.6 Sulfuric Acid -- Concentrated, ACS grade, sp. gr. 1.84.
  - 7.6.1 Sulfuric acid, 12 N -- Slowly add 335 mL concentrated sulfuric acid to 665 mL of reagent water.
- 7.7 Potassium Hydroxide (KOH) -- Pellets, ACS grade.
  - 7.7.1 KOH, 37% (w/v) -- Dissolve 37 g KOH pellets in reagent water and dilute to 100 mL.
- 7.8 Carbitol (Diethylene Glycol Monoethyl Ether) -- ACS grade. Available from Aldrich Chemical Co.
- 7.9 Diazald -- ACS grade. Available from Aldrich Chemical Co.
- 7.10 Diazald Solution -- Prepare a solution containing 10 g Diazald in 100 mL of a 50:50 by volume mixture of ethyl ether and carbitol. This solution is stable for one month or longer when stored at 4°C in an amber bottle with a Teflon-lined screw cap.
- 7.11 Sodium Chloride (NaCl) -- Crystal, ACS Grade. Heat treat in a shallow tray at 450°C for a minimum of four hours to remove interfering organic substances.
- 7.12 4,4'-Dibromooctafluorobiphenyl (DBOB) -- 99% purity, for use as internal standard (available from Aldrich Chemical Co).
- 7.13 2,4-Dichlorophenylacetic Acid (DCAA) -- 99% purity, for use as surrogate standard (available from Aldrich Chemical Co).
- 7.14 Mercuric Chloride -- ACS grade (Aldrich Chemical Co.), for use as a bactericide. If any other bactericide can be shown to work as well as mercuric chloride, it may be used instead.
- 7.15 Reagent Water -- Reagent water is defined as water that is reasonably free of contamination that would prevent the determination of any analyte of interest. Reagent water used to generate the validation data in this method was distilled water obtained from the Magnetic Springs Water Co., Columbus, Ohio.
- 7.16 Silicic Acid -- ACS Grade.
- 7.17 Florisil -- 60-100/PR mesh (Sigma No. F-9127). Activate by heating in a shallow container at 150°C for at least 24 and not more than 48 hours.

- 7.18 Stock Standard Solutions (1.00 µg/µL) -- Stock standard solutions may be purchased as certified solutions or prepared from pure standard materials using the following procedure:
- 7.18.1 Prepare stock standard solutions by accurately weighing approximately 0.0100 g of pure material. Dissolve the material in MTBE and dilute to volume in a 10 mL volumetric flask. Larger volumes may be used at the convenience of the analyst. If compound purity is certified at 96% or greater, the weight may be used without correction to calculate the concentration of the stock standard. Commercially prepared stock standards may be used at any concentration if they are certified by the manufacturer or by an independent source.
  - 7.18.2 Transfer the stock standard solutions into TFE-fluorocarbon-sealed screw cap amber vials. Store at room temperature and protect from light.
  - 7.18.3 Stock standard solutions should be replaced after two months or sooner if comparison with laboratory fortified blanks, or QC samples indicate a problem.
- 7.19 Internal Standard Solution -- Prepare an internal standard solution by accurately weighing approximately 0.0010 g of pure DBOB. Dissolve the DBOB in MTBE and dilute to volume in a 10 mL volumetric flask. Transfer the internal standard solution to a TFE-fluorocarbon-sealed screw cap bottle and store at room temperature. Addition of 25 µL of the internal standard solution to 10 mL of sample extract results in a final internal standard concentration of 0.25 µg/mL. Solution should be replaced when ongoing QC (Section 10.0) indicates a problem. Note that DBOB has been shown to be an effective internal standard for the method analytes<sup>1</sup>, but other compounds may be used if the quality control requirements in Section 10.0 are met.
- 7.20 Surrogate Standard Solution -- Prepare a surrogate standard solution by accurately weighing approximately 0.0010 g of pure DCAA. Dissolve the DCAA in MTBE and dilute to volume in a 10 mL volumetric flask. Transfer the surrogate standard solution to a TFE-fluorocarbon-sealed screw cap bottle and store at room temperature. Addition of 50 µL of the surrogate standard solution to a 1 L sample prior to extraction results in a surrogate standard concentration in the sample of 5 µg/L and, assuming quantitative recovery of DCAA, a surrogate standard concentration in the final extract of 0.5 µg/mL. Solution should be replaced when ongoing QC (Section 10.0) indicates a problem. Note that DCAA has been shown to be an effective surrogate standard for the method analytes<sup>1</sup>, but other compounds may be used if the quality control requirements in Section 10.4 are met.
- 7.21 Laboratory Performance Check Solutions -- Prepare a diluted dinoseb solution by adding 10 µL of the 1.0 µg/µL dinoseb stock solution to the MTBE and diluting to volume in a 10 mL volumetric flask. To prepare the check solution, add 40 µL of the diluted dinoseb solution, 16 µL of the 4-nitrophenol stock solution, 6 µL of

the 3,5-dichlorobenzoic acid stock solution, 50 µL of the surrogate standard solution, 25 µL of the internal standard solution, and 250 µL of methanol to a 5 mL volumetric flask and dilute to volume with MTBE. Methylate sample as described in Section 11.4 or 11.5. Dilute the sample to 10 mL in MTBE. Transfer to a TFE-fluorocarbon-sealed screw cap bottle and store at room temperature. Solution should be replaced when ongoing QC (Section 10.0) indicates a problem.

## **8.0 SAMPLE COLLECTION, PRESERVATION, AND STORAGE**

8.1 Grab samples must be collected in glass containers. Conventional sampling practices<sup>8</sup> should be followed; however, the bottle must not be prerinsed with sample before collection.

### **8.2 Sample Preservation and Storage**

8.2.1 Add mercuric chloride (See Section 7.14) to the sample bottle in amounts to produce a concentration of 10 mg/L. Add 1 mL of a 10 mg/mL solution of mercuric chloride in water to the sample bottle at the sampling site or in the laboratory before shipping to the sampling site. A major disadvantage of mercuric chloride is that it is a highly toxic chemical; mercuric chloride must be handled with caution, and samples containing mercuric chloride must be disposed of properly.

8.2.2 If residual chlorine is present, add 80 mg of sodium thiosulfate per liter of sample to the sample bottle prior to collecting the sample.

8.2.3 After the sample is collected in the bottle containing preservative(s), seal the bottle and shake vigorously for one minute.

8.2.4 The samples must be iced or refrigerated at 4°C away from light from the time of collection until extraction. Preservation study results indicate that the analytes (measured as total acid) present in samples are stable for 14 days when stored under these conditions.<sup>1</sup> However, analyte stability may be affected by the matrix; therefore, the analyst should verify that the preservation technique is applicable to the samples under study.

### **8.3 Extract Storage**

8.3.1 Extracts should be stored at 4°C away from light. Preservation study results indicate that most analytes are stable for 28 days<sup>1</sup>; however, the analyst should verify appropriate extract holding times applicable to the samples under study.

## **9.0 CALIBRATION**

9.1 Establish GC operating parameters equivalent to those indicated in Section 6.10. The GC system may be calibrated using either the internal standard technique (Section 9.2) or the external standard technique (Section 9.3).

NOTE: Calibration standard solutions must be prepared such that no unresolved analytes are mixed together.

9.2 Internal Standard Calibration Procedure -- To use this approach, the analyst must select one or more internal standards compatible in analytical behavior to the compounds of interest. The analyst must further demonstrate that the measurement of the internal standard is not affected by method or matrix interferences. DBOB has been identified as a suitable internal standard.

9.2.1 Prepare calibration standards at a minimum of three (recommend five) concentration levels for each analyte of interest by adding volumes of one or more stock standards to a volumetric flask. To each calibration standard, add a known constant amount of one or more of the internal standards and 250  $\mu\text{L}$  methanol, and dilute to volume with MTBE. Esterify acids with diazomethane as described in Section 11.4 or 11.5. The lowest standard should represent analyte concentrations near, but above, the respective EDLs. The remaining standards should bracket the analyte concentrations expected in the sample extracts, or should define the working range of the detector.

9.2.2 Analyze each calibration standard according to the procedure (Section 11.7). Tabulate response (peak height or area) against concentration for each compound and internal standard. Calculate the response factor (RF) for each analyte and surrogate using Equation 1.

*Equation 1*

$$\text{RF} = \frac{(A_s) (C_{is})}{(A_{is}) (C_s)}$$

where:  $A_s$  = Response for the analyte to be measured.

$A_{is}$  = Response for the internal standard.

$C_{is}$  = Concentration of the internal standard ( $\mu\text{g}/\text{L}$ ).

$C_s$  = Concentration of the analyte to be measured ( $\mu\text{g}/\text{L}$ ).

9.2.3 If the RF value over the working range is constant (20% RSD or less) the average RF can be used for calculations. Alternatively, the results can be used to plot a calibration curve of response ratios ( $A_s/A_{is}$ ) vs.  $C_s$ .

9.2.4 The working calibration curve or RF must be verified on each working shift by the measurement of one or more calibration standards. If the response for any analyte varies from the predicted response by more than  $\pm 20\%$ , the test must be repeated using a fresh calibration standard. If the repetition also fails, a new calibration curve must be generated for that analyte using freshly prepared standards.

- 9.3.5 Single-point calibration is a viable alternative to a calibration curve. Prepare single point standards from the secondary dilution standards in MTBE. The single point standards should be prepared at a concentration that produces a response that deviates from the sample extract response by no more than 20%.
- 9.2.6 Verify calibration standards periodically, recommend at least quarterly, by analyzing a standard prepared from reference material obtained from an independent source. Results from these analyses must be within the limits used to routinely check calibration.

### 9.3 External Standard Calibration Procedure

- 9.3.1 Prepare calibration standards at a minimum of three (recommend five) concentration levels for each analyte of interest and surrogate compound by adding volumes of one or more stock standards and 250  $\mu$ L methanol to a volumetric flask. Dilute to volume with MTBE. Esterify acids with diazomethane as described in Section 11.4 or 11.5. The best standard should represent analyte concentrations near, but above, the respective EDL. The remaining standards should bracket the analyte concentrations expected in the sample extracts, or should define the working range of the detector.
- 9.3.2 Starting with the standard of lowest concentration, analyze each calibration standard according to Section 11.7 and tabulate response (peak height or area) versus the concentration in the standard. The results can be used to prepare a calibration curve for each compound. Alternatively, if the ratio of response to concentration (calibration factor) is a constant over the working range (20% RSD or less), linearity through the origin can be assumed and the average ratio or calibration factor can be used in place of a calibration curve.
- 9.3.3 The working calibration curve or calibration factor must be verified on each working day by the measurement of a minimum of two calibration check standards, one at the beginning and one at the end of the analysis day. These check standards should be at two different concentration levels to verify the calibration curve. For extended periods of analysis (greater than eight hours), it is strongly recommended that check standards be interspersed with samples at regular intervals during the course of the analyses. If the response for any analyte varies from the predicted response by more than  $\pm 20\%$ , the test must be repeated using a fresh calibration standard. If the results still do not agree, generate a new calibration curve or use a single point calibration standard as described in Section 9.3.3.
- 9.3.4 Single-point calibration is a viable alternative to a calibration curve. Prepare single point standards from the secondary dilution standards in MTBE. The single point standards should be prepared at a concentration

that produces a response that deviates from the sample extract response by no more than 20%.

- 9.2.5 Verify calibration standards periodically, recommend at least quarterly, by analyzing a standard prepared from reference material obtained from an independent source. Results from these analyses must be within the limits used to routinely check calibration.

## 10.0 QUALITY CONTROL

- 10.1 Minimum quality control (QC) requirements are initial demonstration of laboratory capability, determination of surrogate compound recoveries in each sample and blank, monitoring internal standard peak area or height in each sample and blank (when internal standard calibration procedures are being employed), analysis of laboratory reagent blanks, laboratory fortified samples, laboratory fortified blanks, and QC samples.
- 10.2 Laboratory Reagent Blanks (LRB) -- Before processing any samples, the analyst must demonstrate that all glassware and reagent interferences are under control. Each time a set of samples is extracted or reagents are changed, a LRB must be analyzed. If within the retention time window of any analyte the LRB produces a peak that would prevent the determination of that analyte, determine the source of contamination and eliminate the interference before processing samples.
- 10.3 Initial Demonstration of Capability
- 10.3.1 Select a representative fortified concentration (about 10 times EDL) for each analyte. Prepare a sample concentrate (in methanol) containing each analyte at 1000 times selected concentration. With a syringe, add 1 mL of the concentrate to each of at least four 1 L aliquots of reagent water, and analyze each aliquot according to procedures beginning in Section 11.0.
- 10.3.2 For each analyte the recovery value for all four of these samples must fall in the range of  $R \pm 30\%$  (or within  $R \pm 3S_R$  if broader) using the values for  $R$  and  $S_R$  for reagent water in Table 2. For those compounds that meet the acceptable criteria, performance is considered acceptable and sample analysis may begin. For those compounds that fail these criteria, this procedure must be reported using five fresh samples until satisfactory performance has been demonstrated.
- 10.3.3 The initial demonstration of capability is used primarily to preclude a laboratory from analyzing unknown samples via a new, unfamiliar method prior to obtaining some experience with it. It is expected that as laboratory personnel gain experience with this method the quality of data will improve beyond those required here.
- 10.4 The analyst is permitted to modify GC columns, GC conditions, detectors, continuous extraction techniques, concentration techniques (i.e., evaporation

techniques), internal standard or surrogate compounds. Each time such method modifications are made, the analyst must repeat the procedures in Section 10.3

## 10.5 Assessing Surrogate Recovery

10.5.1 When surrogate recovery from a sample or method blank is <70% or >130%, check (1) calculations to locate possible errors, (2) spiking solutions for degradation, (3) contamination, and (4) instrument performance. If those steps do not reveal the cause of the problem, reanalyze the extract.

10.5.2 If a blank extract reanalysis fails the 70-130% recovery criterion, the problem must be identified and corrected before continuing.

10.5.3 If sample extract reanalysis meets the surrogate recovery criterion, report only data for the analyzed extract. If sample extract continues to fail the recovery criterion, report all data for that sample as suspect.

## 10.6 Assessing the Internal Standard

10.6.1 When using the internal standard calibration procedure, the analyst is expected to monitor the IS response (peak area or peak height) of all samples during each analysis day. The IS response for any sample chromatogram should not deviate from the daily calibration check standard's IS response by more than 30%.

10.6.2 If >30% deviation occurs with an individual extract, optimize instrument performance and inject a second aliquot of that extract.

10.6.2.1 If the reinjected aliquot produces an acceptable internal standard response, report results for that aliquot.

10.6.2.2 If a deviation of greater than 30% is obtained for the reinjected extract, analysis of the samples should be repeated beginning with Section 11.0, provided the sample is still available. Otherwise, report results obtained from the reinjected extract, but annotate as suspect.

10.6.3 If consecutive samples fail the IS response acceptance criterion, immediately analyze a calibration check standard.

10.6.3.1 If the check standard provides a response factor (RF) within 20% of the predicted value, then follow procedures itemized in Section 10.6.2 for each sample failing the IS response criterion.

10.6.3.2 If the check standard provides a response factor which deviates more than 20% of the predicted value, then the analyst must recalibrate, as specified in Sect. 9.

## 10.7 Assessing Laboratory Performance -- Laboratory Fortified Blank (LFB)

10.7.1 The laboratory must analyze at least one LFB sample with every 20 samples or one per sample set (all samples extracted within a 24-hour period) whichever is greater. The concentration of each analyte in the LFB should be 10 times EDL or the MCL, whichever is less. Calculate accuracy as percent recovery ( $X_p$ ). If the recovery of any analyte falls outside the control limits (see Section 10.7.2), that analyte is judged out of control, and the source of the problem should be identified and resolved before continuing analyses.

10.7.2 Until sufficient data become available from within their own laboratory, usually a minimum of results from 20-30 analyses, the laboratory should assess laboratory performance against the control limits in Section 10.3.2 that are derived from the data in Table 2. When sufficient internal performance data becomes available, develop control limits from the mean percent recovery ( $\bar{X}$ ) and standard deviation (S) of the percent recovery. These data are used to establish upper and lower control limits as follows:

$$\text{UPPER CONTROL LIMIT} = \bar{X} + 3S$$

$$\text{LOWER CONTROL LIMIT} = \bar{X} - 3S$$

After each 5-10 new recovery measurements, new control limits should be calculated using only the most recent 20-30 data points. These calculated control limits should never exceed those established in Section 10.3.2.

10.7.3 It is recommended that the laboratory periodically determine and document its detection limit capabilities for the analytes of interest.

10.7.4 At least quarterly, analyze a QC sample from an outside source.

10.7.5 Laboratories are encouraged to participate in external performance evaluation studies such as the laboratory certification programs offered by many states or the studies conducted by USEPA. Performance evaluation studies serve as independent checks on the analyst's performance.

## 10.8 Assessing Analyte Recovery -- Laboratory Fortified Sample Matrix

10.8.1 The laboratory must add a known concentration to a minimum of 10% of the routine samples or one sample concentration per set, whichever is greater. The concentration should not be less than the background concentration of the sample selected for fortification. Ideally, the concentration should be the same as that used for the laboratory fortified blank (Section 10.7). Over time, samples from all routine sample sources should be fortified.

- 10.8.2 Calculate the percent recovery, P of the concentration for each analyte, after correcting the analytical result, X, from the fortified sample for the background concentration, b, measured in the unfortified sample, i.e.,:

$$P = 100 (X - b) / \text{fortifying concentration},$$

and compare these values to control limits appropriate for reagent water data collected in the same fashion. If the analyzed unfortified sample is found to contain **NO** background concentrations, and the added concentrations are those specified in Section 10.7, then the appropriate control limits would be the acceptance limits in Section 10.7. If, on the other hand, the analyzed unfortified sample is found to contain background concentration, b, estimate the standard deviation at the background concentration,  $s_b$ , using regressions or comparable background data and, similarly, estimate the mean,  $\bar{X}_a$  and standard deviation,  $s_a$ , of analytical results at the total concentration after fortifying. Then the appropriate percentage control limits would be  $\bar{P} \pm 3s_p$ , where:

$$\bar{P} = 100 \bar{X} / (b + \text{fortifying concentration})$$

$$\text{and } s_p = 100 (s_a^2 + s_b^2)^{1/2} / \text{fortifying concentration}$$

For example, if the background concentration for Analyte A was found to be 1 µg/L and the added amount was also 1 µg/L, and upon analysis the laboratory fortified sample measured 1.6 µ/L, then the calculated P for this sample would be (1.6 µg/L minus 1.0 µg/L)/ 1 µg/L or 60%. This calculated P is compared to control limits derived from prior reagent water data. Assume it is known that analysis of an interference free sample at 1 µg/L yields an s of 0.12 µg/L and similar analysis at 2.0 µg/L yields  $\bar{X}$  and s of 2.01 µg/L and 0.20 µg/L, respectively. The appropriate limits to judge the reasonableness of the percent recovery, 60%, obtained on the fortified matrix sample is computed as follows:

$$\begin{aligned} & [100 (2.01 \mu\text{g/L}) / 2.0 \mu\text{g/L}] \\ & \pm 3 (100) [(0.12 \mu\text{g/L})^2 + (0.20 \mu\text{g/L})^2]^{1/2} / 1.0 \mu\text{g/L} = \\ & 100.5\% \pm 300 (0.233) = \\ & 100.5\% \pm 70\% \text{ or } 30\% \text{ to } 170 \text{ recovery of the added analyte.} \end{aligned}$$

- 10.8.3 If the recovery of any such analyte falls outside the designated range, and the laboratory performance for that analyte is shown to be in control (Section 10.7), the recovery problem encountered with the fortified sample is judged to be matrix related, not system related. The result for that analyte in the unfortified sample is labeled suspect/matrix to inform the data user that the results are suspect due to matrix effects.

- 10.9 Assessing Instrument System -- Laboratory Performance Check Sample (LPC) - Instrument performance should be monitored on a daily basis by analysis of the LPC sample. The LPC sample contains compounds designed to indicate appropriate instrument sensitivity, column performance (primary column) and chromatographic performance. LPC sample components and performance criteria are listed in Table 3. Inability to demonstrate acceptable instrument performance indicates the need for reevaluation of the instrument system. The sensitivity requirements are set based on the EDLs published in this method. If laboratory EDLs differ from those listed in this method, concentrations of the instrument QC standard compounds must be adjusted to be compatible with the laboratory EDLs.
- 10.10 The laboratory may adopt additional quality control practices for use with this method. The specific practices that are most productive depend upon the needs of the laboratory and the nature of the samples. For example, field or laboratory duplicates may be analyzed to assess the precision of the environmental measurements or field reagent blanks may be used to assess contamination of samples under site conditions, transportation and storage.

## 11.0 **PROCEDURE**

### 11.1 Manual Hydrolysis, Preparation, and Extraction

- 11.1.1 Add preservative to blanks and QC check standards. Mark the water meniscus on the side of the sample bottle for later determination of sample volume (Section 11.1.9). Pour the entire sample into a 2 L separatory funnel. Fortify sample with 50  $\mu$ L of the surrogate standard solution.
- 11.1.2 Add 250 g NaCl to the sample, seal, and shake to dissolve salt.
- 11.1.3 Add 17 mL of 6 N NaOH to the sample, seal, and shake. Check the pH of the sample with pH paper; if the sample does not have a pH greater than or equal to 12, adjust the pH by adding more 6 N NaOH. Let the sample sit at room temperature for one hour, shaking the separatory funnel and contents periodically.
- 11.1.4 Add 60 mL methylene chloride to the sample bottle to rinse the bottle, transfer the methylene chloride to the separatory funnel and extract the sample by vigorously shaking the funnel for two minutes with periodic venting to release excess pressure. Allow the organic layer to separate from the water phase for a minimum of 10 minutes. If the emulsion interface between layers is more than one-third the volume of the solvent layer, the analyst must employ mechanical techniques to complete the phase separation. The optimum technique depends upon the sample, but may include stirring, filtration through glass wool, centrifugation, or other physical methods. Discard the methylene chloride phase.

- 11.1.5 Add a second 60 mL volume of methylene chloride to the sample bottle and repeat the extraction procedure a second time, discarding the methylene chloride layer. Perform a third extraction in the same manner.
- 11.1.6 Add 17 mL of 12 N H<sub>2</sub>SO<sub>4</sub> to the sample, seal, and shake to mix. Check the pH of the sample with pH paper; if the sample does not have a pH less than or equal to 2, adjust the pH by adding more 12 N H<sub>2</sub>SO<sub>4</sub>.
- 11.1.7 Add 120 mL ethyl ether to the sample, seal, and extract the sample by vigorously shaking the funnel for two minutes with periodic venting to release excess pressure. Allow the organic layer to separate from the water phase for a minimum of 10 minutes. If the emulsion interface between layers is more than one-third the volume of the solvent layer, the analyst must employ mechanical techniques to complete the phase separation. The optimum technique depends upon the sample, but may include stirring, filtration through glass wool, centrifugation, or other physical methods. Remove the aqueous phase to a 2 L Erlenmeyer flask and collect the ethyl ether phase in a 500 mL round-bottom flask containing approximately 10 g of acidified anhydrous sodium sulfate. Periodically, vigorously shake the sample and drying agent. Allow the extract to remain in contact with the sodium sulfate for approximately two hours.
- 11.1.8 Return the aqueous phase to the separatory funnel, add a 60 mL volume of ethyl ether to the sample, and repeat the extraction procedure a second time, combining the extracts in the 500 mL erlenmeyer flask. Perform a third extraction with 60 mL of ethyl ether in the same manner.
- 11.1.9 Determine the original sample volume by refilling the sample bottle to the mark and transferring the water to a 1000 mL graduated cylinder. Record the sample volume to the nearest 5 mL.
- 11.2 Automated Hydrolysis, Preparation, and Extraction -- Data presented in this method were generated using the automated extraction procedure with the mechanical separatory funnel shaker.
- 11.2.1 Add preservative (Section 8.2) to any samples not previously preserved, e.g., blanks and QC check standards. Mark the water meniscus on the side of the sample bottle for later determination of sample volume (Section 11.2.9). Fortify sample with 50 µL of the surrogate standard solution. If the mechanical separatory funnel shaker is used, pour the entire sample into a 2 L separatory funnel. If the mechanical tumbler is used, pour the entire sample into a tumbler bottle.
- 11.2.2 Add 250 g NaCl to the sample, seal, and shake to dissolve salt.
- 11.2.3 Add 17 mL of 6 N NaOH to the sample, seal, and shake. Check the pH of the sample with pH paper; if the sample does not have a pH greater

than or equal to 12, adjust the pH by adding more 6 N NaOH. Shake sample for one hour using the appropriate mechanical mixing device.

- 11.2.4 Add 300 mL methylene chloride to the sample bottle to rinse the bottle, transfer the methylene chloride to the separatory funnel or tumbler bottle, seal, and shake for 10 seconds, venting periodically. Repeat shaking and venting until pressure release is not observed during venting. Reseal and place sample container in appropriate mechanical mixing device. Shake or tumble the sample for one hour. Complete and thorough mixing of the organic and aqueous phases should be observed at least two minutes after starting the mixing device.
- 11.2.5 Remove the sample container from the mixing device. If the tumbler is used, pour contents of tumbler bottle into a 2 L separatory funnel. Allow the organic layer to separate from the water phase for a minimum of 10 minutes. If the emulsion interface between layers is more than one third the volume of the solvent layer, the analyst must employ mechanical techniques to complete the phase separation. The optimum technique depends upon the sample, but may include stirring, filtration through glass wool, centrifugation, or other physical methods. Drain and discard the organic phase. If the tumbler is used, return the aqueous phase to the tumbler bottle.
- 11.2.6 Add 17 mL of 12 N H<sub>2</sub>SO<sub>4</sub> to the sample, seal, and shake to mix. Check the pH of the sample with pH paper; if the sample does not have a pH less than or equal to 2, adjust the pH by adding more 12 N H<sub>2</sub>SO<sub>4</sub>.
- 11.2.7 Add 300 mL ethyl ether to the sample, seal, and shake for 10 seconds, venting periodically. Repeat shaking and venting until pressure release is not observed during venting. Reseal and place sample container in appropriate mechanical mixing device. Shake or tumble sample for one hour. Complete and thorough mixing of the organic and aqueous phases should be observed at least two minutes after starting the mixing device.
- 11.2.8 Remove the sample container from the mixing device. If the tumbler is used, pour contents of tumbler bottle into a 2 L separatory funnel. Allow the organic layer to separate from the water phase for a minimum of 10 minutes. If the emulsion interface between layers is more than one-third the volume of the solvent layer, the analyst must employ mechanical techniques to complete the phase separation. The optimum technique depends upon the sample, but may include stirring, filtration through glass wool, centrifugation, or other physical methods. Drain and discard the aqueous phase. Collect the extract in a 500-mL round-bottom flask containing about 10 g of acidified anhydrous sodium sulfate. Periodically vigorously shake the sample and drying agent. Allow the extract to remain in contact with the sodium sulfate for approximately two hours.

- 11.2.9 Determine the original sample volume by refilling the sample bottle to the mark and transferring the water to a 1000 mL graduated cylinder. Record the sample volume to the nearest 5 mL.
- 11.3 Extract Concentration
- 11.3.1 Assemble a K-D concentrator by attaching a concentrator tube to a 500 mL evaporative flask.
- 11.3.2 Pour the dried extract through a funnel plugged with acid washed glass wool, and collect the extract in the K-D concentrator. Use a glass rod to crush any caked sodium sulfate during the transfer. Rinse the round-bottom flask and funnel with 20-30 mL of ethyl ether to complete the quantitative transfer.
- 11.3.3 Add one to two clean boiling stones to the evaporative flask and attach a macro Snyder column. Prewet the Snyder column by adding about 1 mL ethyl ether to the top. Place the K-D apparatus on a hot water bath, 60-65°C, so that the concentrator tube is partially immersed in the hot water, and the entire lower rounded surface of the flask is bathed with hot vapor. At the proper rate of distillation the balls of the column will actively chatter but the chambers will not flood. When the apparent volume of liquid reaches 1 mL, remove the K-D apparatus and allow it to drain and cool for at least 10 minutes.
- 11.3.4 Remove the Snyder column and rinse the flask and its lower joint into the concentrator tube with 1-2 mL of ethyl ether. Add 2 mL of MTBE and a fresh boiling stone. Attach a micro-Snyder column to the concentrator tube and prewet the column by adding about 0.5 mL of ethyl ether to the top. Place the micro K-D apparatus on the water bath so that the concentrator tube is partially immersed in the hot water. Adjust the vertical position of the apparatus and the water temperature as required to complete concentration in 5-10 minutes. When the apparent volume of liquid reaches 0.5 mL, remove the micro K-D from the bath and allow it to drain and cool. Remove the micro Snyder column and add 250 µL of methanol. If the gaseous diazomethane procedure (Section 11.4) is used for esterification of pesticides, rinse the walls of the concentrator tube while adjusting the volume to 5.0 mL with MTBE. If the pesticides will be esterified using the diazomethane solution (Section 11.5), rinse the walls of the concentrator tube while adjusting the volume to 4.5 mL with MTBE.
- 11.4 Esterification of Acids Using Gaseous Diazomethane -- Results presented in this method were generated using the gaseous diazomethane derivatization procedure. See Section 11.5 for an alternative procedure.
- 11.4.1 Assemble the diazomethane generator (Figure 1) in a hood.

- 11.4.2 Add 5 mL of ethyl ether to Tube 1. Add 1 mL of ethyl ether, 1 mL of carbitol, 1.5 mL of 37% aqueous KOH, and 0.2 grams Diazald to Tube 2. Immediately place the exit tube into the concentrator tube containing the sample extract. Apply nitrogen flow (10 mL/min) to bubble diazomethane through the extract for one minute. Remove first sample. Rinse the tip of the diazomethane generator with ethyl ether after methylation of each sample. Bubble diazomethane through the second sample extract for one minute. Diazomethane reaction mixture should be used to esterify only two samples; prepare new reaction mixture in Tube 2 to esterify each two additional samples. Samples should turn yellow after addition of diazomethane and remain yellow for at least two minutes. Repeat methylation procedure if necessary.
- 11.4.3 Seal concentrator tubes with stoppers. Store at room temperature in a hood for 30 minutes.
- 11.4.4 Destroy any unreacted diazomethane by adding 0.1-0.2 g silicic acid to the concentrator tubes. Allow to stand until the evolution of nitrogen gas has stopped (approximately 20 minutes). Adjust the sample volume to 5.0 mL with MTBE.
- 11.5 Esterification of Acids Using Diazomethane Solution -- Alternative procedure
- 11.5.1 Assemble the diazomethane generator (Figure 2) in a hood. The collection vessel is a 10 mL or 15 mL vial, equipped with a Teflon-lined screw cap and maintained at 0-5°C.
- 11.5.2 Add a sufficient amount of ethyl ether to Tube 1 to cover the first impinger. Add 5 mL of MTBE to the collection vial. Set the nitrogen flow at 5-10 mL/min. Add 2 mL Diazald solution (Section 7.10) and 1.5 mL of 37% KOH solution to the second impinger. Connect the tubing as shown and allow the nitrogen flow to purge the diazomethane from the reaction vessel into the collection vial for 30 minutes. Cap the vial when collection is complete and maintain at 0-5°C. When stored at 0-5°C this diazomethane solution may be used over a period of 48 hours.
- 11.5.3 To each concentrator tube containing sample or standard, add 0.5 mL diazomethane solution. Samples should turn yellow after addition of the diazomethane solution and remain yellow for at least two minutes. Repeat methylation procedure if necessary.
- 11.5.4 Seal concentrator tubes with stoppers. Store at room temperature in a hood for 30 minutes.
- 11.5.5 Destroy any unreacted diazomethane by adding 0.1-0.2 g silicic acid to the concentrator tubes. Allow to stand until the evolution of nitrogen gas has stopped (approximately 20 minutes). Adjust the sample volume to 5.0 mL with MTBE.

## 11.6 Florisil Separation

- 11.6.1 Place a small plug of glass wool into a 5 mL disposable glass pipet. Tare the pipet, and measure 1 g of activated Florisil into the pipet.
- 11.6.2 Apply 5 mL of 5% methanol in MTBE to the Florisil. Allow the liquid to just reach the top of the Florisil. In this and subsequent steps, allow the liquid level to just reach the top of the Florisil before applying the next rinse, however, do not allow the Florisil to go dry. Discard eluate.
- 11.6.3 Apply 5 mL methylated sample to the Florisil leaving silicic acid in the tube. Collect eluate in K-D tube.
- 11.6.4 Add 1 mL of 5% methanol in MTBE to the sample container, rinsing walls. Transfer the rinse to the Florisil column leaving silicic acid in the tube. Collect eluate in a K-D tube. Repeat with 1 mL and 3 mL aliquots of 5% methanol in MTBE, collecting eluates in K-D tube.
- 11.6.5 If necessary, dilute eluate to 10 mL with 5% methanol in MTBE.
- 11.6.6 Seal the vial and store in a refrigerator if further processing will not be performed immediately. Analyze by GC-ECD.

## 11.7 Gas Chromatography

- 11.7.1 Section 6.10 summarizes the recommended operating conditions for the GC. Included in Table 1 are retention times observed using this method. Other GC columns, chromatographic conditions, or detectors may be used if the requirements of Section 10.4 are met.
- 11.7.2 Calibrate the system daily as described in Section 9.0. The standards and extracts must be in MTBE.
- 11.7.3 If the internal standard calibration procedure is used, fortify the extract with 25  $\mu\text{L}$  of internal standard solution. Thoroughly mix sample and place aliquot in a GC vial for subsequent analysis.
- 11.7.4 Inject 2  $\mu\text{L}$  of the sample extract. Record the resulting peak size in area units.
- 11.7.5 If the response for the peak exceeds the working range of the system, dilute the extract and reanalyze.

## 11.8 Identification of Analytes

- 11.8.1 Identify a sample component by comparison of its retention time to the retention time of a reference chromatogram. If the retention time of an

unknown compound corresponds, within limits, to the retention time of a standard compound, then identification is considered positive.

- 11.8.2 The width of the retention time window used to make identifications should be based upon measurements of actual retention time variations of standards over the course of a day. Three times the standard deviation of a retention time can be used to calculate a suggested window size for a compound. However, the experience of the analyst should weigh heavily in the interpretation of chromatograms.
- 11.8.3 Identification requires expert judgement when sample components are not resolved chromatographically. When GC peaks obviously represent more than one sample component (i.e., broadened peak with shoulder(s) or valley between two or more maxima, or any time doubt exists over the identification of a peak on a chromatogram, appropriate alternative techniques, to help confirm peak identification, need to be employed. For example, more positive identification may be made by the use of an alternative detector which operates on a chemical/physical principle different from that originally used, e.g., mass spectrometry, or the use of a second chromatography column. A suggested alternative column is described in Section 6.10.

## 12.0 CALCULATIONS

- 12.1 Calculate analyte concentrations in the sample from the response for the analyte using the calibration procedure described in Section 9.0.
- 12.2 If the internal standard calibration procedure is used, calculate the concentration (C) in the sample using the response factor (RF) determined in Section 9.2 and Equation 2, or determine sample concentration from the calibration curve.

*Equation 2*

$$C (\mu\text{g/L}) = \frac{(A_s) (I_s)}{(A_{is}) (RF) (V_o)}$$

where:  $A_s$  = Response for the parameter to be measured.

$A_{is}$  = Response for the internal standard.

$I_s$  = Amount of internal standard added to each extract ( $\mu\text{g}$ ).

$V_o$  = Volume of water extracted (L).

- 12.3 If the external standard calibration procedure is used, calculate the amount of material injected from the peak response using the calibration curve or calibration factor determined in Section 9.3. The concentration (C) in the sample can be calculated from Equation 3.

*Equation 3*

$$C (\mu\text{g/L}) = \frac{(A) (V_i)}{(V_t) (V_s)}$$

where: A = Amount of material injected (ng).

$V_i$  = Volume of extract injected ( $\mu\text{L}$ ).

$V_t$  = Volume of total extract ( $\mu\text{L}$ ).

$V_s$  = Volume of water extracted (mL).

### 13.0 PRECISION AND ACCURACY

13.1 In a single laboratory, analyte recoveries from reagent water were determined at five concentration levels. Results were used to determine analyte EDLs and demonstrate method range.<sup>1</sup> Analyte EDLs and analyte recoveries and standard deviation about the percent recoveries at one concentration are given in Table 2.

13.2 In a single laboratory, analyte recoveries from one standard synthetic ground waters were determined at one concentration level. Results were used to demonstrate applicability of the method to different ground water matrices.<sup>1</sup> Analyte recoveries from the one synthetic matrix are given in Table 2.

### 14.0 REFERENCES

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**TABLE 1. RETENTION TIMES FOR METHOD ANALYTES**

Analyte	Retention Time <sup>a</sup> (minutes)	
	Primary	Confirmation
Dalapon	3.4	4.7
3,5-Dichlorobenzoic acid	18.6	17.7
4-Nitrophenol	18.6	20.5
DCAA (surrogate)	22.0	14.9
Dicamba	22.1	22.6
Dichlorprop	25.0	25.6
2,4-D	25.5	27.0
DBOB (int. std.)	27.5	27.6
Pentachlorophenol (PCP)	28.3	27.0
Chloramben	29.7	32.8
2,4,5-TP	29.7	29.5
5-Hydroxydicamba	30.0	30.7
2,4,5-T	30.5	30.9
2,4-DB	32.2	32.2
Dinoseb	32.4	34.1
Bentazon	33.3	34.6
Picloram	34.4	37.5
DCCA acid metabolites	<sup>3</sup> 5.8	37.8
Acifluorfen	41.5	42.8

<sup>a</sup>Columns and analytical conditions are described in Sections 6.10.1 and 6.10.2.

**TABLE 2. SINGLE LABORATORY ACCURACY, PRECISION AND ESTIMATED DETECTION LIMITS (EDLS) FOR ANALYTES FROM REAGENT WATER AND SYNTHETIC GROUNDWATERS<sup>a</sup>**

Analyte	EDL µg/L <sup>b</sup>	Concentration µg/L	Reagent Water		Synthetic Water 1 <sup>e</sup>	
			R <sup>c</sup>	S <sub>r</sub> <sup>d</sup>	R	S <sub>R</sub>
Acifluorfen	0.096	0.2	121	15.7	103	20.6
Bentazon	0.2	1	120	16.8	82	37.7
Chloramben	0.093	0.4	111	14.4	112	10.1
2,4-D	0.2	1	131	27.5	110	5.5
Dalapon	1.3	10	100	20.0	128	30.7
2,4-DB	0.8	4	87	13.1	0	0
DCPA acid metabolites	0.02	0.2	74	9.7	81	21.9
Dicamba	0.081	0.4	135	32.4	92	17.5
3,5-Dichlorobenzoic acid	0.061	0.6	102	16.3	82	7.4
Dichlorprop	0.26	2	107	20.3	106	5.3
Dinoseb	0.19	0.4	42	14.3	89	13.4
5-Hydroxydicamba	0.04	0.2	103	16.5	88	5.3
4-Nitrophenol	0.13	1	131	23.6	127	34.3
Pentachlorophenol (PCP)	0.076	0.04	130	31.2	84	9.2
Picloram	0.14	0.6	91	15.5	97	23.3
2,4,5-T	0.08	0.4	117	16.4	96	3.8
2,4,5-TP	0.075	0.2	134	30.8	105	6.3

<sup>a</sup>Data corrected for amount detected in blank and represent the mean of seven to eight samples.

<sup>b</sup>EDL = estimated detection limit; defined as either MDL (Appendix B to 40 CFR Part 136 - Definition and Procedure for the Determination of the Method Detection Limit - Revision 1.11) or a level of compound in a sample yielding a peak in the final extract with signal-to-noise ratio of approximately 5, whichever value is higher. The concentration used in determining the EDL is not the same as the concentration presented in this table.

<sup>c</sup>R = average percent recovery.

<sup>d</sup>S<sub>r</sub> = standard deviation of the percent recovery

<sup>e</sup>Corrected for amount found in blank; Absopure Nature Artesian Spring Water Obtained from the Absopure Water Company in Plymouth, Michigan.

**TABLE 3. LABORATORY PERFORMANCE CHECK SOLUTION**

<b>Test</b>	<b>Analyte</b>	<b>Conc, µg/mL</b>	<b>Requirements</b>
Sensitivity	Dinoseb	0.004	Detection of analyte; S/N >3
Chromatographic performance	4-Nitrophenol	1.6	0.70 < PGF <1.05
Column performance	3,5-Dichlorobenzoic acid	0.6	Resolution >0.40 <sup>b</sup>
	4-Nitrophenol	1.6	

<sup>a</sup>PGF - peak Gaussian factor. Calculated using the equation:

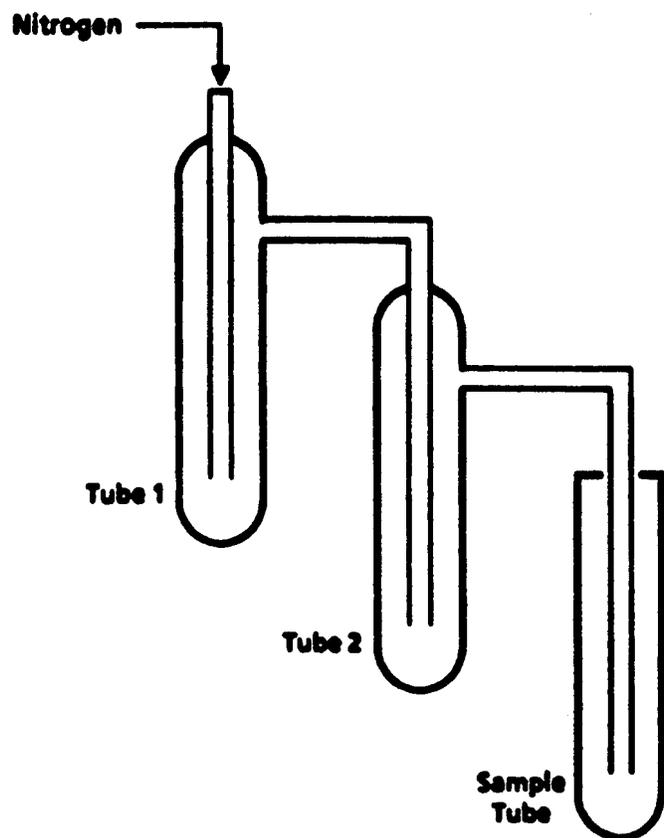
$$PGF = \frac{1.83 \times W(1/2)}{W(1/10)}$$

where: W(1/2) is the peak width at half height and W(1/10) is the peak width at tenth height.

<sup>b</sup>Resolution between the two peaks as defined by the equation:

$$R = \frac{t}{W}$$

where: t is the difference in elution times between the two peaks and W is the average peak width, at the baseline, of the two peaks.



**FIGURE 1. GASEOUS DIAZOMETHANE GENERATOR**

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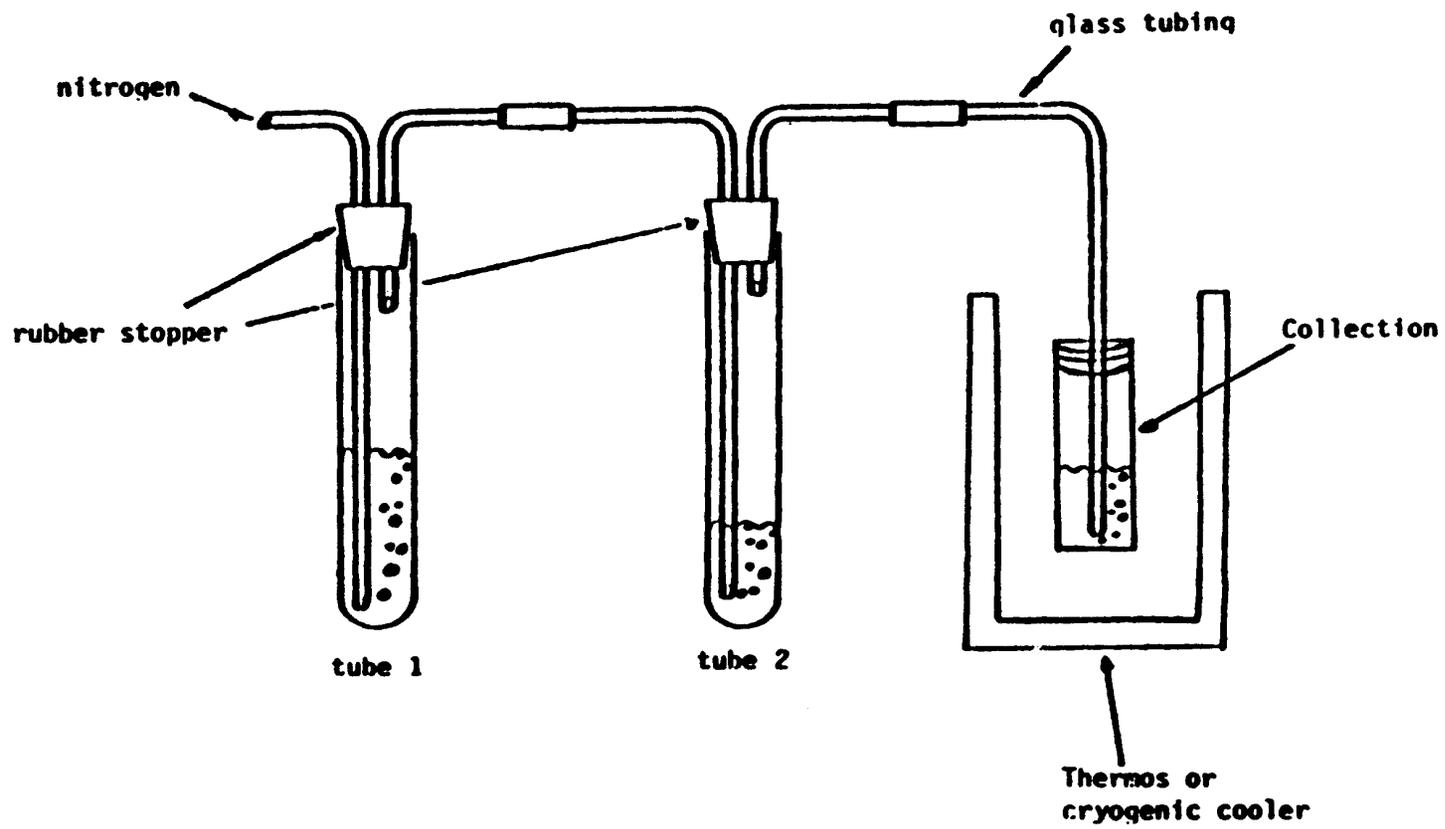


FIGURE 2. DIAZOMETHANE SOLUTION GENERATOR